

CORN HUSK FERMENTATION WITH BREWERY WASTE YEAST AS FISH MEAL SUBSTITUTE IN THE DIET OF AFRICAN CATFISH (*Clarias gariepinus*) FINGERLINGS

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ABSTRACT

This study was conducted to evaluate the effect of partial substitution of fish-meal (FM) with two types of fermented animal by-products (fish and poultry offal's wastes) on growth performance, microbiological, and fish quality of African catfish (*Clarias gariepinus*). Fish offal's silage (FOS) and poultry offal's silage (POS) were fermented by *Lactobacillus acidophilus*. Five isonitrogenous (34.3% crude protein) diets were formulated to comprise two levels of fermented fish offal (20% and 40%), two levels of fermented poultry offal (20% and 40%) wastes, and 100% fish meal as a control. Ninety hatcheries bred African catfish were randomly distributed into 5 equal groups every 18 fish in three replicate (6 fish/ replicate aquaria). Each of the diets was fed to triplicate groups of African catfish with mean initial weight (MIW) $52\text{ g} \pm 0.01$ at 5% body weight in glass aquaria (30 x 30 x70 cm) twice daily (morning and evening) for 12 weeks. The results showed that fish in the fourth group fed on diet supplemented with 20 % POS recorded the best feed conversion ratio (FCR) (1.77 ± 0.1), protein efficiency ratio (PER) (3.51 ± 0.06), incidence of cost (IC) (16.3 ± 0.37), and profit index (PI) (3.51 ± 0.06). Fish in group four as well had the least total bacterial count, coliform, and mold & yeast. The least lactic acid bacteria count and the best overall acceptance were detected also in the fourth group as compared to other groups except diet 1(control) which means it can be used safely without significantly affecting public health or consumer acceptance.

Keywords: Fish-meal replacement, Growth performance, Cost efficiency, Microbial count, organoleptic parameters.

INTRODUCTION

Fish meal (FM) has been used in aquafeeds for decades due to its high protein content, balanced amino acid profile, high digestibility, palatability and as a source of essential fatty acids. However, FM has currently become expensive as well as scarce due to its extensive use in animal and fish feed industries (Akegbejo-Samsons and Fasakin, 2008). Therefore, researchers are looking for other alternative protein sources that can lower the level of FM used in fish feed formulation without significantly affecting growth performance and fish quality and yet profitable. Two promising animal by-products used for replacement of FM are fish offal and poultry offal due to their high protein content, high level of lipid which may cause a protein-sparing effect, low

carbohydrate content, and lack of anti-nutritional factors, but the use of animal by-products was also constrained by high moisture, indigestible particles, microbial contaminants, etc. (Cruz-Suárez *et al.*, 2007; Samaddar and Kaviraj, 2014). Fermentation is an effective biotechnological tool that can be used to overcome many of the before mentioned problems associated with the utilization of animal by-products in fish diets that can reduce the cost of aquafeeds and protect the environment from microbial contamination related to the disposal of animal wastes. Fermentation also improved the nutritional quality and digestibility of the ingredients (Vidotti *et al.*, 2003). Lactic acid bacteria (LAB) were widely employed in fermentation for animal feed production (Driehuis *et al.*, 2001, Samaddar and Kaviraj, 2014 and Zhao *et al.*, 2019). Moreover, it is a simpler, faster, eco-friendlier, and economical process (Ahmed and Mahendrakar, 1996, Raghunath and Gopakumar, 2002).

Therefore, the current study was conducted to investigate the effect of partial replacement of FM with fermented fish fermented offal's silage and

poultry offal's silage on growth performance, microbiological and organoleptic fish quality of African catfish (*Clarias gariepinus*).

MATERIALS AND METHODS

1. Experimental fish

Ninety hatcheries bred African catfish (*Clarias gariepinus*) (mean initial weight 52 ± 0.01 g) were obtained from the Central laboratory for aquaculture research (CLAR) at Abbassa, Abou-Hammad, Sharkia-Egypt. Fish were transported to the laboratory at Nutrition and Clinical Nutrition Department, Faculty of Veterinary Medicine, Suez Canal University. Upon arrival, all fish were treated with a solution of 3% NaCl for 15 min to eliminate ectoparasites infection, acclimated to the experimental conditions for one week and fed the control diet (100% FM). Hatchery bred African catfish were randomly distributed into 5 groups each of 18 fish in three replicates (6 fish/ replicate aquaria).

2. Fermentation

Samples of fish offal (comprising mainly heads and intestine of tilapia fishes and mullets) and samples of poultry offal were collected from the local fish market in Ismailia province, transported in ice buckets to avoid further contamination to the laboratory and stored at -20°C before processing. Cane molasses also, collected from the local market was used as a source of fermentable carbohydrate for microorganisms involved in the fermentation.

Five kilograms of fish offal and poultry offal samples each were weighed and thoroughly rinsed in tap water before blended into a paste. Fifty ml/kg of actively growing culture of *Lactobacillus acidophilus* and 150g / kg of cane molasses were added to the paste and allowed to ferment for 14 days in airtight plastic containers. The pH of the silage dropped from the initial value of 7.4 to 4.12 on the 14th day from the start of the fermentation process. The desired level of pH indicative of completion of fermentation is 4.0 – 4.2 indicates that the fermented products are safe from spoilage and fit for use as a feedstuff for animals (Lee *et al.*, 2003; Hashoda *et al.*, 2001). The liquid product was then oven-dried at 60°C for 48 h. The dried product was ground into fine powder to form the silage meal, and stored at -20°C .

3. Feed formulation

Five isonitrogenous (34.3 % crude protein) diets were formulated by replacing dietary fish meal at 20%, 40% with fermented fish offal's silage, 20%, 40% fermented poultry offal's silage and control diet (100% fish meal) was also formulated. The ingredients of each diet were mixed thoroughly at the required ratios (Table 1) in Braun mixing machine to obtain a homogeneous mass. Corn starch and hot

water were added and mixed further to obtain a dough-like paste. The diets mixtures were then passed through a 2-mm die (Braun) mincing machine to form model-like strands that were air-dried and stored at -20°C in air-tight polyethylene bags. The strands were mechanically broken before feeding into pellets of a suitable size for African catfish (*Clarias gariepinus*).

4. Experimental design

At the start of the experiment, 15 glass aquaria (40 X 45 X 80 Cm) were each stocked with 6 fish with an average weight of 52 ± 0.01 g. Each diet was assigned to triplicate groups of fish. Fish were fed at a rate of 5% of its body weight, divided into two equal feedings (morning and evening). Fish from each aquarium were weighed collectively every two weeks and the amount of diet fed was adjusted accordingly.

5. Sampling and analysis

Proximate analysis

Proximate analysis of random samples of fish offal's silage and poultry offal silage was carried out according to AOAC, 1990. Data obtained were used in the feed formulation of experimental diets as shown in Table 2.

Growth performance

At the end of the feeding trial, fish were starved for 24 hours, counted and weighed. The obtained data were used in growth performance calculations:

Total body weight gain = Final body weight (g) – Initial body weight (g)

Feed conversion ratio FCR = feed intake (g)/ body weight gain (g)

Protein efficiency ratio PER = Protein intake (g)/ body weight gain (g)

Economic analysis

Economic analysis was carried out to estimate the cost of feed to raise a kilogram of fish using the experimental diets:

Incidence of cost IC = cost of feed/weight gain

Profit index PI = Value of fish /Cost of feed

Microbiological

Three random samples of fish from each experimental group were taken. Ten grams of each sample were mixed with 90 ml of sterile buffered 0.1 % peptone water in a sterile blender, under sterile conditions, to give 1/10 dilution. Serial dilutions were prepared to be used for counting total bacteria count, total coliform count, Salmonella, Lactobacillus, Staphylococci, yeast, and mold counts according to the procedures described by (Manual, 1984).

Organoleptic assessment

Three random samples of fish from each experimental group were filleted fillets rinsed with cold water then 2 fillets (average weight 20 g) were cut from the central portion, placed in individual covered foil dishes and held at 5°C until removed to room temp 24°C to equilibrate for 1 h. before cooking. Samples were placed on trays, cooked in an electric oven at 200°C for 6 min. then transferred to a holding oven at 75°C for up to half an hour before tasting. Ten panelists were evaluated for taste, odor, texture, appearance and overall acceptability. The assessment was based on the scoring system by Minim, (2006) which involved the measurement of the previously mentioned parameters on a 7-point hedonic scale for the determination of selected characteristics. The

seven categories were ranked as follows: very much liked (7), liked a lot (6), liked (5) liked and didn't liked (4), disliked (3), much disliked (2) and very much disliked (1).

6. Statistical analysis

Data are presented as treatment means \pm S. E. The data were analyzed by one-way ANOVA. When ANOVA identified differences among groups, multiple comparisons among means were made with Duncan's multiple ranges tests. Statistical significance was determined by setting the aggregate type I error at 5% ($P < 0.05$) for each set of comparisons. All the calculations were carried out using the statistical package SPSS version 14 for windows.

Table 1 Proximate composition of fermented fish offal's silage and poultry offal's silage (air-dried sample)

Nutrient (%)	fish offal's silage	poultry offal's silage
Moisture	76.4 \pm 0.8	66.3 \pm 0.4
Crude Protein	35.2 \pm 0.7	41.7 \pm 0.7
Ether Extract	7.9 \pm 1.3	24.8 \pm 1.1
Ash	8.6 \pm 0.2	6.3 \pm 0.3

Table 2: Dietary composition of experimental diets according to recommendations of (NRC, 2011).

Ingredient%	Diet 1 (control)	Diet 2 (20% FOS)	Diet 3 (40% FOS)	Diet 4 (20% POS)	Diet 5 (40% POS)
Poultry meal (66%)	19.4	19.7	20.3	19.5	20
Fish meal (61.6%)	10	8	6	8	6
Poultry offal's silage (41.7%)	0.0	0.0	0.0	2	4
Fish offal's silage (35.2%)	0.0	2	4	0.0	0.0
Ground Y. Corn (8.7%)	21.1	19.8	18.8	19.7	19
Soybean meal (41.6%)	26.5	27.6	28.1	27.7	28
Wheat middling (17.1%)	15	14.6	14.5	15	15
# Oil mix	6	6.1	6.1	6.1	6
*Premix (Vit. & Min.)	0.3	0.3	0.3	0.3	0.3
Common salt	0.3	0.3	0.3	0.3	0.3
Mono calcium phosphate	1.4	1.6	1.6	1.4	1.4
Crude protein (%)	34.3 \pm 0.7	34.3 \pm 0.8	34.3 \pm 0.4	34.3 \pm 1.3	34.3 \pm 1.6
Digestible energy	2950 Kcal DE/ kg	2950 Kcal DE/ kg	2950 Kcal DE/ kg	2950 Kcal DE/ kg	2950 Kcal DE/ kg

*Premix, each 3 kg contains the following vitamins, minerals, monocalcium phosphate, binder and salt: "Vit.A 15 MIU, Vit. D3 2 MIU, Vit. E 1,000 mg, Vit. k3 1,000 mg, Vit. B11,000 mg, Vit. B2 5,000 mg, Vit. B6 1,500 mg, Vit. B12 10 mg, biotin 50 mg, pantothenic acid 10,000 mg, nicotinic acid 30,000 mg, folic acid 1,000 mg, manganese 60,000 mg, zinc 50,000 mg, iron 30,000 mg, copper 4,000 mg, iodine 300 mg, selenium 100 mg, cobalt 100 mg", carrier (CaCO₃) to 3 kg (Golden premix, Egypt).

linseed oil plus olein (1:1).

The analysis was performed according to the Official Methods of Analytical Association of Official Analytical Chemists, Arlington (AOAC, 1990)

RESULTS

Growth performance data (Table 3 and Figure 1) showed that there were significant differences ($P < 0.05$) between groups in the final average weight of fish (FW) and weight gain (WG) at the end of the 12 weeks feeding trial. Fish in the fifth group (40% POS) recorded the highest FW and WG compared to other groups but lower than the control one, while both 20% POS and 20% FOS supplemented groups recorded the same value. Fish in 40% FOS supplemented group recorded the lowest value. There were significant differences ($P < 0.05$) between experimental groups in feed conversion ratio (FCR) where fish in fifth and third groups recorded the worst values compared to the control group and the other treated groups. The best values were recorded in both the control and fourth (POS 20%) groups.

Regarding the cost evaluation data in Table 4, the profit index (IP) obtained by using the diets was highest in the third group (40% FOS) as compared to the control group which recorded the lowest value. The highest incidence of the cost was found in the fifth group (40% POS) compared with the control group which was the lowest.

Microbiological data (Table 5) showed that at the end of the feeding trial, there were significant differences ($P < 0.05$) in total bacterial count between groups where FOS 40% supplemented group recorded the highest value followed by the control group and POS 40% group, while POS 20% and FOS 20% groups recorded the lowest value. Almost the same results were obtained in coliform bacteria. There were no significant differences between groups in Staphylococcus. Salmonella was not detected in all 5 groups. For Mold and Yeast results, FOS 40% and POS 20% recorded the highest value followed by POS 40% while both POS 20% and the control group recorded the lowest value.

Organoleptic parameter data (Table 6) showed that there were no significant differences ($P < 0.05$) in color between all groups except FOS 40% group which recorded a lower value compared to other groups. Significant differences ($P < 0.05$) between groups in taste and texture were detected where the control and POW 20% recorded the highest value followed by FOW 20% group while both FOW 40% and POW 40% groups recorded the lowest value. There were significant differences ($P < 0.05$) between groups also in odor.

Table 3: Growth performance parameters of African catfish, *Clarias gariepinus* fed the experimental diets (means \pm S.E).

Parameter	Diet 1 (control)	Diet 2 (20% FOS)	Diet 3 (40% FOS)	Diet 4 (20% POS)	Diet 5 (40% POS)
Initial Wt. (g)	299.7 \pm 1.3	304.7 \pm 4.2	303.0 \pm 1.1	301.0 \pm 1.3	299.7 \pm 1.6
Final Wt. (g)	440.3 \pm 4.6 ^a	389.7 \pm 1.6 ^c	374.7 \pm 1.3 ^d	385.3 \pm 3.6 ^c	410.7 \pm 3.1 ^b
Weight Gain (g)	140.6 \pm 3.3 ^a	85.0 \pm 4 ^c	64.6 \pm 1.3 ^d	84.3 \pm 7.2 ^c	111.0 \pm 2.1 ^b
Feed intake (g)	245.8 \pm 2.7 ^a	167.2 \pm 7 ^b	136.3 \pm 1.3 ^b	149.2 \pm 6.5 ^b	246.9 \pm 8 ^a
FCR	1.75 \pm 0.1 ^{cd}	2.0 \pm 0.1 ^{bc}	2.1 \pm 0.1 ^{ab}	1.77 \pm 0.1 ^{cd}	2.2 \pm 0.1 ^{ab}
PER	1.67 \pm 0.01 ^a	1.48 \pm 0.00 ^{bc}	1.38 \pm 0.00 ^c	1.58 \pm 0.03 ^{ab}	1.24 \pm 0.06 ^d

^{abc} means in the same row with different superscripts are significantly different ($P < 0.05$).

FCR: feed conversion ratio, PER: Protein efficiency ratio.

Table 4: Costs evaluation of experimental diets fed to African catfish, *Clarias gariepinus* (means \pm S.E).

Value	Diet 1 (control)	Diet 2 (20% FOS)	Diet 3 (40% FOS)	Diet 4 (20% POS)	Diet 5 (40% POS)
Feed consumed (g)	245.8 \pm 2.7 ^a	167.2 \pm 7 ^b	136.3 \pm 1.3 ^b	149.2 \pm 6.5 ^b	246.9 \pm 8 ^a
Weight gain (g)	140.6 \pm 3.3 ^a	85.0 \pm 4 ^c	64.6 \pm 1.3 ^d	84.3 \pm 7.2 ^c	111.0 \pm 2.1 ^b
Cost of feed (LE)	2.26 \pm 0.01 ^a	1.48 \pm 0.03 ^b	1.15 \pm 0.01 ^d	1.37 \pm 0.03 ^c	2.22 \pm 0.09 ^a
Value of fish (LE)	5.28 \pm 0.02 ^a	4.76 \pm 0.01 ^c	4.49 \pm 0.01 ^d	4.62 \pm 0.03 ^c	4.92 \pm 0.06 ^b
Incidence of cost *	16.08 \pm 0.12 ^c	17.43 \pm 0.05 ^{bc}	17.86 \pm 0.08 ^b	16.30 \pm 0.37 ^{bc}	19.79 \pm 1.04 ^a
Profit index**	2.43 \pm 0.00 ^d	3.16 \pm 0.06 ^c	3.89 \pm 0.01 ^a	3.51 \pm 0.06 ^b	2.23 \pm 0.09 ^d

^{abcd} means in the same row with different superscripts are significantly different ($P < 0.05$).

*: Incidence of cost = cost of feed / Kg fish produced, **: Profit index = value of fish / cost of feed

Table 5: Microbial population of African catfish, *Clarias gariepinus* fed the experimental diets (means \pm S.E)

Microorganism (Log CFU/ml)	Diet 1 (control)	Diet 2 (20% FOS)	Diet 3 (40% FOS)	Diet 4 (20% POS)	Diet 5 (40% POS)
Total bacterial count	4.82 \pm 0.27 ^{bc}	4.27 \pm 0.18 ^c	5.44 \pm 0.35 ^b	4.12 \pm 0.18 ^c	5.03 \pm 0.39 ^{bc}
Coliform	3.36 \pm 0.32 ^{bc}	3.65 \pm 0.10 ^{abc}	4.13 \pm 0.17 ^a	2.95 \pm 0.27 ^c	3.98 \pm 0.20 ^{ab}
Salmonella	ND	ND	ND	ND	ND
Staphylococci	4.36 \pm 0.27	4.11 \pm 0.12	4.31 \pm 0.19	4.00 \pm 0.16	4.27 \pm 0.30
Mold & yeast	4.35 \pm 0.26 ^{bc}	5.06 \pm 0.20 ^a	5.47 \pm 0.16 ^a	3.76 \pm 0.23 ^c	4.86 \pm 0.22 ^{bc}
LAB	1.71 \pm 0.11 ^c	3.28 \pm 0.35 ^{bc}	3.71 \pm 0.32 ^a	2.39 \pm 0.29 ^{bc}	3.11 \pm 0.31 ^{bc}

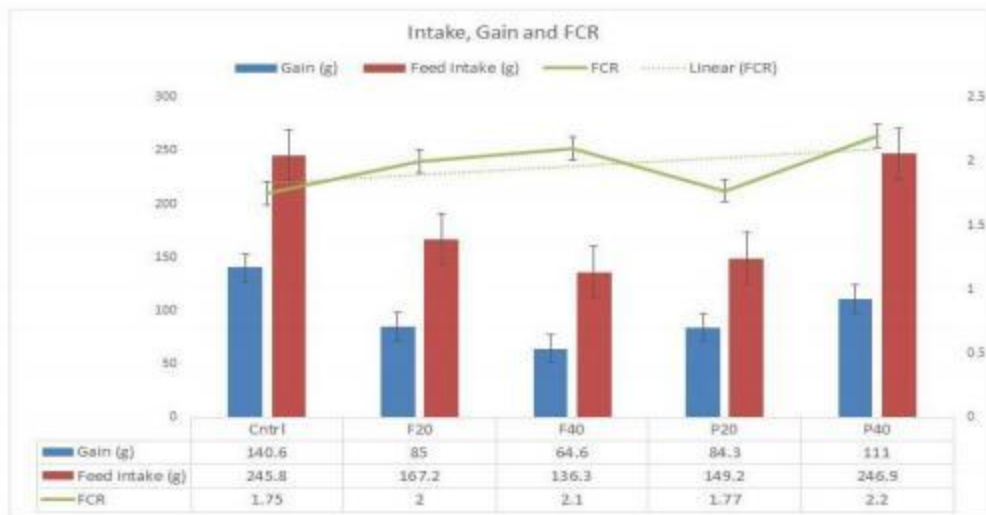
^{abc} means in the same row with different superscripts are significantly different (P<0.05).

LAB: Lactic acid bacteria, ND: Not detected.

Table 6: Organoleptic parameters of African catfish, *Clarias gariepinus* fed the experimental diets (means \pm S.E)

Physical parameters	Diet 1 (control)	Diet 2 (20% FOS)	Diet 3 (40% FOS)	Diet 4 (20% POS)	Diet 5 (40% POS)
Taste	6.66 \pm 0.33 ^a	5.83 \pm 0.16 ^{ab}	5.00 \pm 0.57 ^b	6.33 \pm 0.33 ^a	5.66 \pm 0.33 ^b
Odour	6.83 \pm 0.16 ^a	5.00 \pm 0.57 ^{bc}	4.00 \pm 0.57 ^c	6.33 \pm 0.66 ^{ab}	5.56 \pm 0.33 ^{ab}
Texture	6.33 \pm 0.33 ^a	5.00 \pm 0.57 ^{ab}	4.00 \pm 0.57 ^b	6.83 \pm 0.18 ^a	5.46 \pm 0.33 ^b
Appearance	6.83 \pm 0.16 ^a	5.66 \pm 0.33 ^b	4.33 \pm 0.33 ^c	6.83 \pm 0.16 ^a	5.00 \pm 0.57 ^{bc}
Color	6.50 \pm 0.28 ^a	5.33 \pm 0.33 ^a	4.00 \pm 0.57 ^b	6.00 \pm 0.28 ^a	5.33 \pm 0.33 ^a
Overall acceptability	6.64 \pm 0.09 ^a	5.42 \pm 0.12 ^b	4.33 \pm 0.33 ^c	6.44 \pm 0.24 ^{ab}	5.38 \pm 0.05 ^b

^{abc} means in the same row with different superscripts are significantly different (P<0.05).

**Figure 1** Feed intake, body weight gain and feed conversion ratio (FCR) relationship of African catfish fed the experimental diets.

DISCUSSION

The proximate composition of bacterially fermented poultry offal's silage and fish offal's silage used in experimental diets formulation is shown in Table 1. Crude protein percent of poultry offal's silage (41.7 ± 0.7) and fish offal's silage (35.2 ± 0.7) showed that silage is rich in protein and compares favorably with other conventional dietary protein sources. Similar results were obtained by Soltan and Tharwat, (2006) who reported 32.51% CP in fermented fish silage and Belal *et al.* (1995) who reported 39.5% CP in chicken offal's silage.

Data of growth performance parameters (Table 3, Fig. 1) and production costs evaluation (Table 4) revealed that there were significant differences ($P < 0.05$) between groups in FW, WG, FI, FCR and PER were fish in control group recorded the best growth and feed utilization parameters and the least profit index (PI). The high digestibility, palatability and the balanced amino acid profile of FM and its high price also, could explain the better growth performance and feed utilization of the control group, yet remains costly. Fish in the fourth group (20% POS) recorded the highest FCR and PER and the best IC and PI as compared to the other treated groups except for the control group. Fermentation breaks down protein which facilitates nutrient utilization as fish can assimilate protein as amino acids and short peptides, hence the better performance (FCR, 1.77 and PER, 1.58) and consequently, the better IC and PI of diet 4 together with the level of inclusion (20% POS). Similar results were obtained by Higgs (1979) who reported that at least 28% of poultry by-product meal may be included in the diet of Coho salmon, without amino acid supplementation and Tacon (1993) who reported that poultry by-products showed similarities with fishmeal based on the nutritional composition. Also, Steffens (1994) reported that poultry by-product meal is suitable as a partial or complete replacement, but that complete substitution required amino acid supplementation, principally with lysine and methionine. This could explain why diet 5 recorded the highest FW and WG compared to other groups but less than diet 1 and also the highest FI similar to diet 1. However, it wasn't successful when it comes to feeding utilization FCR, IC, PI and PER which were the least among all experimental groups. Also, Fagbenro and Bello-Olusoji (1997) reported that poultry offal can replace up to 40% of a high-quality fishmeal protein without amino acid supplementation, whilst not compromising growth performance and feed utilization.

In contrast, Soltan and Tharwat (2006) reported a non-significant effect on growth and feed utilization parameters and reduced feed costs /kg diet and feed costs /kg weight gain by 15.59 and 19.39%, respectively when a fish meal was replaced by 50% fermented fish silage. Our data showed that diet 2

(20% FOS) recorded lower FCR, PER, IC, PI values than diet 4 (20% POS) while, Higher FCR, PER and IC values than diet 3 (40% FOS). Moreover, Mondal *et al.* (2008) reported that fermented fish offal could replace 30% of FM in diets of *Labeo rohita* and *Heteropneustes fossilis*.

The purpose of microbial population tests is to detect pathogenic bacteria (*Salmonella*, *Staphylococcus aureus*, *E. coli*) or indicator organisms of fecal pollution (fecal coliforms, fecal streptococci) or other types of general contamination or poor handling practices (coliform bacteria, faecal streptococci, total viable count). Organoleptic assessment is a sensory assessment using senses to judge the quality of a product. Both parameters are important in the aquaculture industry to ensure food safety and consumer acceptance.

Microbial population and organoleptic parameters of African catfish, *Clarias gariepinus* fed the experimental diets are shown in Table 5 and Table 6. Statistical analysis of the obtained data revealed that there were significant differences ($P < 0.05$) between groups in TBC, Coliform, Mold & yeast and LAB where diet 4 recorded the least TBC, coliform and mold & yeast as compared to all other groups, while the least LAB and overall acceptance as compared to the other groups except the control group. There were no significant differences between groups in staphylococcus count and salmonella was not detected in any of the groups. However, the highest degree of hazardous microorganisms (TBC, Coliform, Mold & Yeast and LAB) was found in diet 3 (40% FOS). The presence of these microorganisms in fish is of great concern to public health. Fish in the third group also recorded the lowest overall acceptance which might be quite understandable taking into consideration its high degree of contamination.

CONCLUSION

Based on nutrient utilization indices, estimated economic benefits, microbial count and overall acceptance, poultry offal's silage can effectively replace fishmeal in African catfish (*Clarias gariepinus*) diets till the level of 20% without affecting growth, microbial and organoleptic qualities for a profitable and sustainable aquaculture venture.

CONFLICTS OF INTEREST

The authors declare that there is no competing interest.

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تأثيرات استبدال مسحوق السمه بمخلفات حيواويت متخمرة في علائك لراميط شمال افريقيا علي النمو ومحتوى الجسم مه العاصر الغذائية وجودة الاسمان

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لجزء ١٠٠ من أدراسة حم ١١ جأت از الاسحدذاي أزشي سخق أسه بعا ١٠٠ إحزات أح ١٠٠ اة أخرة
(فوات خفات الأسان ١٠٠ أداج) ع أداء ١٠٠ ، أماسات الأربا ١٠٠ جاة ، خدة
أسان أمراط. ج جاز س الاج خفات الأسان س الاج فضلات أداج بأسطة بحز ١٠٠ الأوباس اس اس انفاس. ج جى .
خس علايك حش حراس (٣٪ ٣٤ ١٠٠ أدزج ١٠٠ أخا) . شخ ع سخ ١٠٠ ١٠٠ خفات الأسان ٢٠) ٪ ٤٠ .
٪ ١٠٠ حح ١٠٠ سخق أسه (وذه ١٠٠ سخ ١٠٠ ١٠٠ خفات أداج أخرة ٢٠) ٪ ٤٠ ٪ ١٠٠
حح ١٠٠ سخق أسه (ع ١٠٠ ضابط ١٠٠ ١٠٠ إسحدذاي سخق أسه بخفات الأسان ١٠٠ أداج أخر
١٠٠) ٪ ١٠٠ سزق أسه. ج اسخذأ عدد ٩٠ ١٠٠ أسان أمراط ج جمس ١٠٠ ١٠٠ خس زعات و
زغ با 3 بزرات ج جعد ١٠٠ الأسان ع أعلايك زج ١٠٠ ل ١٠٠ بعدي ٥ ٪ ١٠٠ سن أزس. سزث
أز ع أزابع أجد ع ١٠٠ با ٢٠ ٪ خفات أداج سدة ج ١٠٠ لأعلاف) ١٠٠ ، ٧٧ ، ٠ ، ١٠٠ ، ٠٠ سدة وفاء
أدزج ١٠٠ (٥٠ ، ٣ ± ٦٠ ، ٠) ، ١٠٠ ، ٠٠ (٥٠ ، ٣ ± ٦٠ ، ٠) ، ١٠٠ ، ٠٠ ع ١٠٠
الاسعاضة ع ٢٠ ٪ ١٠٠ جدات الأسان بس الاج أداج أخرة ف عف سه أمراط أخاس و أضح
أزرزب؛ ١٠٠ اج س ١٠٠ اسخذأ عف ، جخفاض جفاف أعضة أضي ع أزبح. ١٠٠ ضا أجد ف
أز ع أزابع جذا ١٠٠ أل عدد ببحاز و ، ل ١٠٠ ، أفضوات أخ ازة. أل عدد ببحاز ١٠٠ الأوحاه
اساذ أفض لذي عا ١٠٠ ضا مارة بازعات الأخر باسحباء إطا أذاي ف أز ع الإ ١٠٠ (أبحزي)
١٠٠ ، ١٠٠ اسخذأ ب ١٠٠ د ١٠٠ ألتاز بشى وداز ع ١٠٠ أصحة أعلة أ لذي أسح ١٠٠ .

